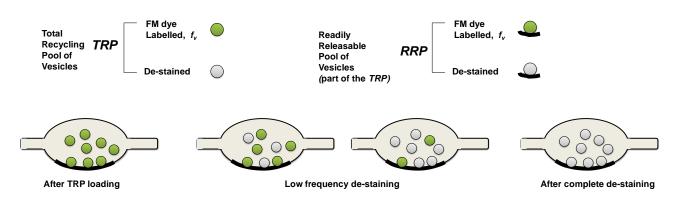
Text S1. Quantification of FM dye (SRC1) de-staining during low frequency stimulation.

In this section we consider a quantitative model which allows estimating basic presynaptic parameters using SRC1 de-staining profiles recorded in individual synaptic

boutons.



Notation:

- f_{v} specific fluorescence of a single SRC1 labeled vesicle
- RRP number of readily-releasable vesicles
- TRP total number of recycling vesicles
- p_v average release probability of individual vesicles within RRP

 $F_{FM}(t)$ specific vesicular FM dye fluorescence intensity in individual synaptic boutons

Model assumptions:

In general, FM dye de-staining profiles have complex shapes that depend on the type of FM dye used and on loading/stimulation protocols [1–5]. It has however been shown that vesicles within RRP and TRP co-exist in a dynamic equilibrium [6,7]. Therefore we hypothesized that, during low frequency (0.5 Hz) stimulation, there is enough time for

newly endocytosed fluorescence-free vesicles to re-equilibrate between the TRP and RRP between successive APs. To test this hypothesis we compared SRC1 de-staining at 0.5 Hz and 0.25 Hz. If SRC1 de-staining was significantly affected by depletion of RRP vesicles or by preferential recycling of RPP vesicles (for example as has been reported after bursts of high frequency stimulation [7]), then the SRC1 de-staining per AP should be higher at 0.25 Hz than at 0.5 Hz. However, no such difference between SRC1 de-staining rates at these two frequencies was found when k_{AP} was normalized by the number of APs (Figure S1 G - I). We therefore conclude that, during low frequency stimulation, fluorescence-labeled and fluorescence-free vesicles were equally distributed between RRP and TRP.

We then obtain:

$$\frac{d_{FM}F(t)}{dt} = -\nu \cdot N_{AP} \cdot f_{\nu} \cdot \frac{F_{FM}(t)}{\Delta F_{FMtotal}}$$
(1.1)

where $\Delta F_{FMtotal} = TRP \cdot f_v$ is the specific fluorescence of FM dye labeled vesicles corresponding to completely labeled TRP, and N_{AP} is the average number of vesicles released during a single AP. Under the binomial model [8–10], the average number of vesicles released during a single AP (i.e. vesicular release rate) follows the relationship: $N_{AP} = RRP \cdot p_v$. This yields:

$$\frac{dF_{FM}(t)}{dt} = -v \cdot [RRP/TRP] \cdot p_v \cdot F_{FM}(t)$$
 and

$$F_{FM}(t) = \Delta F_{FMtotal} \cdot \exp(-\nu \cdot [RRP/TRP] \cdot p_{\nu} \cdot t)$$

Thus the experimentally determined AP-specific SRC1 de-staining rate can be expressed as:

$$k_{AP} = v \cdot \left[RRP / TRP \right] p_{v} \tag{1.2}$$

Consequently, another experimentally determined parameter - the vesicular release rate

 $(R_{rel} = \frac{\Delta F_{FMtotal}k_{AP}}{v})$ – is proportional to the average number of vesicles released during a

single AP - $R_{rel} \propto N_{AP}$:

$$R_{rel} = \frac{\Delta F_{FMtotal} k_{AP}}{V} = TRP \cdot f_v \cdot [RRP / TRP] \cdot p_v = f_v \cdot RRP \cdot p_v = f_v \cdot N_{AP}$$

Reference List

- 1. Klingauf J, Kavalali ET, Tsien RW (1998) Kinetics and regulation of fast endocytosis at hippocampal synapses. Nature 394: 581-585.
- Vanden BP, Klingauf J (2006) Synaptic vesicles in rat hippocampal boutons recycle to different pools in a use-dependent fashion. J Physiol 572: 707-720.
- 3. Mozhayeva MG, Sara Y, Liu X, Kavalali ET (2002) Development of vesicle pools during maturation of hippocampal synapses. J Neurosci 22: 654-665.
- 4. Murthy VN, Sejnowski TJ, Stevens CF (1997) Heterogeneous release properties of visualized individual hippocampal synapses. Neuron 18: 599-612.
- Waters J, Smith SJ (2002) Vesicle pool partitioning influences presynaptic diversity and weighting in rat hippocampal synapses. J Physiol 541: 811-823.
- 6. Murthy VN, Stevens CF (1999) Reversal of synaptic vesicle docking at central synapses. Nat Neurosci 2: 503-507.
- 7. Pyle JL, Kavalali ET, Piedras-Renteria ES, Tsien RW (2000) Rapid reuse of readily releasable pool vesicles at hippocampal synapses. Neuron 28: 221-231.
- 8. Schneggenburger R, Sakaba T, Neher E (2002) Vesicle pools and short-term synaptic depression: lessons from a large synapse. Trends Neurosci 25: 206-212.
- 9. Christie JM, Jahr CE (2006) Multivesicular release at Schaffer collateral-CA1 hippocampal synapses. J Neurosci 26: 210-216.
- 10. Oertner TG, Sabatini BL, Nimchinsky EA, Svoboda K (2002) Facilitation at single synapses probed with optical quantal analysis. Nat Neurosci 5: 657-664.