PROTOCOL S1: CALCULATION OF THE $m_{salt}$ SLOPES FROM THE EXPERIMENTAL $T_m$ VALUES.

For the wild type forms of Hv 1ALigN, Ec 1ALigN and ProtL the curves of protein stability versus temperature have been estimated from equilibrium denaturation experiments (using guanidinium chloride) and thermal denaturation experiments (Figure A).

**Figure A**: Protein stability versus temperature for wild type ProtL (red), (previously reported in (1)), wild type Hv 1ALigN (blue) and wild type Ec 1ALigN (green). Circles correspond to the experimental $\Delta G^U$-$F$ points whereas solid lines show the best fitting to the Gibbs-Helmholtz equation.

The thermodynamic parameters have been determined by fitting the experimental data to the Gibbs-Helmholtz curve:

**Table A**: Thermodynamic parameters for WT ProtL, WT Hv 1ALigN and WT Ec 1ALigN.

<table>
<thead>
<tr>
<th>Parameter</th>
<th>WT ProtL</th>
<th>WT Hv 1ALigN</th>
<th>WT Ec 1ALigN</th>
</tr>
</thead>
<tbody>
<tr>
<td>$\Delta H_m$ / kcal·mol$^{-1}$</td>
<td>$53.0 \pm 4.8^{(1,2)}$</td>
<td>$27.0 \pm 2.5^{(1)}$</td>
<td>$28.9 \pm 2.1^{(1)}$</td>
</tr>
<tr>
<td>$\Delta C_p$ / kcal·mol$^{-1}$·K$^{-1}$</td>
<td>$0.876 \pm 0.05^{(1)}$</td>
<td>$0.461 \pm 0.02^{(1)}$</td>
<td>$0.650 \pm 0.1^{(1)}$</td>
</tr>
</tbody>
</table>

(1) The error has been estimated by Montecarlo propagation of the individual experimental errors, obtained from duplicates.

(2) Differential scanning calorimetry data for wild type ProtL gives a value of 54.5 kcal·mol$^{-1}$
Free energy changes upon salt addition can be calculated (for a given protein and with respect to a reference state) from the experimental $T_m$ values (2):

$$
\Delta G_{salt} = \Delta H_{salt} \left[ 1 - \frac{T_m^0}{T_{salt}} \right] - \Delta C_p^* \left[ (T_{salt}^{salt} - T_m^0) + T_m^0 \ln \left( \frac{T_m^0}{T_{salt}^{salt}} \right) \right]
$$

where $T_{salt}$ and $\Delta H_{salt}$ are the melting temperature and melting enthalpy in the presence of salt respectively and $\Delta C_p^*$ is the apparent heat capacity change in the presence of cosolute. The label "salt" accounts for KCl or NaCl and $T_m^0$ is the melting temperature at the reference state (specified in Figure A for Hv 1ALigN, Ec 1ALigN and ProtL).

$\Delta C_p^*$ can be estimated from the slope of a plot of $\Delta H_m$ versus $T_m$ (2). For each mutant and at every salt concentration, $T_m$ and $\Delta H_m$ values have been obtained from the fitting of the thermal denaturation curves. Plots of $\Delta H_m$ versus $T_m$ result in flat lines with negligible slopes. Figure B shows the plot of $\Delta H_m$ versus their respective $T_m$ for all the mutants and conditions studied.

**Figure B:** $\Delta H_m$ versus $T_m$ for the ProtL (red circles), Hv 1ALigN (blue circles) and Ec 1ALigN (green circles) datasets. All the mutants and each salt concentration tested in the present study are included in the plot. Values for wild type proteins (solid lines) and their confidence limits (dashed lines) are also represented. For a subset of data, $\Delta H_m$ have also been determined from DSC data (white circles).
\( \Delta H_m \) values are always very close to the ones for the reference state (represented by solid lines in Figure B), almost always falling within the error limits for the reference state (shown as dashed lines in Figure B). For a selected set of data, the \( \Delta H_m \) values have also been obtained from differential scanning calorimetry data (see below). The calorimetric \( \Delta H_m \) values show less scattering from the reference enthalpies. All together it can be concluded that \( \Delta C_p^{\text{ProtL}} = \Delta C_p^{\text{Hv 1ALigN}} = \Delta C_p^{\text{Ec 1ALigN}} = 0 \) and the free energy changes upon salt addition have been determined from the equation:

\[
\Delta G_{\text{salt}} = \Delta H_m^0 \left( 1 - \frac{T_0}{T_{\text{salt}}} \right)
\]

where \( \Delta H_m^0 \) is equal to 53, 28.9 and 27 kcal·mol\(^{-1}\) for ProtL, \( Ec 1\text{ALigN} \) and \( Hv 1\text{ALigN} \) respectively and \( T_0 \) is the reference state for the three proteins (ProtL = 20 mM phosphate, pH 6.0; \( Hv 1\text{ALigN} = 20 \) mM phosphate, pH 8.0, 1M KCl; \( Ec 1\text{ALigN} = 20 \) mM phosphate, pH 8.0).

\( \Delta G_{\text{salt}} \) varies linearly with respect to the molar salt concentration and the slope \( (m_{\text{salt}}) \) is used as a metric for haloadaptation:

\[
m_{\text{salt}} = \left( \frac{\partial \Delta G_{\text{salt}}}{\partial C_m} \right)
\]

where \( C_m \) is the molar salt concentration.

**Estimation of the error.** Error in \( m_{\text{salt}} \) has been propagated from the experimental uncertainties of the \( T_m \) by montecarlo analysis: for each \( T_m \), 10000 random variants have been generated within the limits of the experimental uncertainty and they have been transformed into \( \Delta G_{\text{salt}} \). This dataset has been used to generate an ensemble of \( m_{\text{salt}} \) values. The standard deviation of this distribution is used as the error estimation for \( m_{\text{salt}} \).

**DSC data.** For differential scanning calorimetry (DSC) measurements, sample concentration oscillated between 0.4 mM and 1.6 mM. For a given buffer condition, calorimeter was pre-equilibrated overnight and samples were run from 15 °C to 95 °C at 1 °C/min. Data analysis included cubic baseline-progression and two state model data fitting with the apparatus-associated software based in Origin (MicroCal). Data shown in Figure B corresponds to ProtL: wild type (no salt, 500 mM NaCl and 1M NaCl), Kx1Q (no salt), Kx2Q
(no salt), Kx3Q (no salt and 1M NaCl), Kx5Q (2M NaCl), Kx5E (2M NaCl) and Ex6D (1M NaCl) and 1ALigN: wild type (2M KCl), Ex4Q (2M and 3M KCl) where "no salt" means 20 mM phosphate buffer at pH 6.0.